Evaluation of Quantum Energy Field Effects on Cellular ATP Production Rates in Human Cell Lines

Authors:
Robert Sheaff, PhD

Abstract

This study investigated whether quantum field exposure generated by Leela Quantum Tech technology influences adenosine triphosphate (ATP) production in human cell lines. This double-blind experiment was performed using A549 human lung carcinoma cells and immortalized human diploid fibroblasts (HDFs). ATP was quantified using the CellTiter-Glo assay following remote quantum field treatment applied from > 500 miles away. Progressive refinements in the experimental design minimized variability related to cell attachment, incubation conditions, and medium composition. Across the experiment, treated cells exhibited transient but statistically significant elevations in ATP production relative to matched controls, with increases ranging from 20 to 29 percent. The findings suggest that non-local quantum field exposure may modulate cellular bioenergetics. Additional replication and mechanistic work are required to confirm these observations and elucidate underlying pathways.

Introduction

Adenosine triphosphate (ATP) serves as the universal energy currency of the cell, driving nearly all anabolic reactions, signal transduction pathways, and mechanical work within living systems (1, 2). ATP production reflects the integrated activity of mitochondrial oxidative phosphorylation and cytosolic glycolysis, and even minor variations in its synthesis or turnover can profoundly affect cellular physiology, including growth, differentiation, and responses to stress (3–5). Because ATP is continually utilized and replenished, its steady-state levels provide a sensitive indicator of the cell's overall bioenergetic balance.

A range of environmental and physical factors—including temperature shifts, oxidative stress, electromagnetic fields, and photonic influences—have been reported to modulate ATP synthesis or mitochondrial function (7). These findings have prompted growing interest in whether other energetic interactions might also alter bioenergetic processes.

Leela Quantum Tech has developed a proprietary technology designed to concentrate and project quantum energy fields capable of interacting with biological systems. The theoretical model of quantum entanglement proposes that two or more systems can maintain correlations even when physically separated, allowing non-local interactions to occur without classical energy transfer (9). Within this theoretical framework, it becomes a testable question whether such non-local quantum field exposure can produce measurable effects on living cells.

Materials and Methods

Two human cell lines were used to investigate the effects of quantum field exposure on cellular adenosine triphosphate (ATP) production.

A549 cells, a human non-small-cell lung carcinoma line, served as a metabolically active epithelial model, while human diploid fibroblasts (HDFs) immortalized by telomerase expression represented a non-transformed cell type with standard metabolic characteristics.

Cells were cultured in Dulbecco's Modified Eagle Medium (DMEM; Gibco) supplemented with 10% fetal bovine serum (FBS), 100 U/mL penicillin, and 100 µg/mL streptomycin. Cultures were maintained at 37 °C in a humidified incubator with 5% CO_2 unless otherwise specified.

Design

An independent, double-blind experiment was conducted to evaluate the potential influence of a proprietary quantum field (Leela Quantum Tech) on cellular ATP production. In the experiment, two identically prepared groups of cells were randomly labeled "A" and "B." One group was exposed to the quantum field ("treated"), while the other served as a sham control ("untreated"). The exposure procedure was coordinated remotely by Leela Quantum Tech personnel located more than 500 miles away.

To preserve experimental integrity, neither the experimenter (R. Sheaff) nor laboratory staff performing measurements were aware of which group was treated until all data were collected and recorded. Group assignments were revealed only after data analysis was complete.

Measurements were conducted in quadruplicate, and standard deviations were calculated for each condition. Statistical analysis was performed using a one-way analysis of variance (ANOVA) to assess differences between treated and control samples across time points. A significance threshold of $p < 0.05$ was applied. The report indicates that statistically significant increases in ATP levels were observed in treated cells.

Results

A549 and HDF cells were distributed into two separate 96-well plates specifically designed to allow cell attachment. The use of attachment plates ensured proper adherence of both cell types prior to analysis. To prevent fluctuations in pH that could occur with repeated removal of plates from a CO_2 incubator, the experiment was performed at 37 °C in the absence of CO_2. For these conditions, L15 medium with or without glucose was used, as it is formulated for culture under non-CO_2 conditions.

Following plating, cells were incubated for seven hours at 37 °C to allow attachment, confirmed by microscopy, but not long enough for cell replication. After incubation, the plates were randomly labeled “A” and “B” and transferred to a colleague who performed the quantum exposure procedure in collaboration with Leela Quantum Tech personnel located more than 500 miles away. One plate was exposed to the quantum field (“treated”), and the other served as the control (“untreated”). This stage of the experiment was performed entirely in the absence of the principal investigator to preserve a double-blind design. Within five minutes, both plates were returned to the incubator, and at predefined time points, ATP levels were analyzed using the CellTiter-Glo® (CTG) luminescent assay applied directly to each well. Measurements were performed in quadruplicate at each time point to improve data quality and statistical reliability.

A transient but **statistically significant increase** in ATP levels was observed in the treated cells compared with the untreated controls. This increase occurred in both the presence and absence of glucose, indicating that the observed effect was **not restricted to a specific metabolic pathway**. In A549 cells, ATP levels increased by approximately **20 – 29 %**, while HDF cells also showed a transient elevation in ATP concentration, particularly in the glucose-containing condition.

Although the signal in HDFs was less pronounced than in A549 cells, the trend toward higher ATP production in treated samples was clearly evident. The removal of the “spill-over” effect—previously identified as possible cross-influence between charged

and uncharged plates—likely contributed to the stronger response observed in this optimized configuration, where untreated cells remained entirely unexposed to the field.

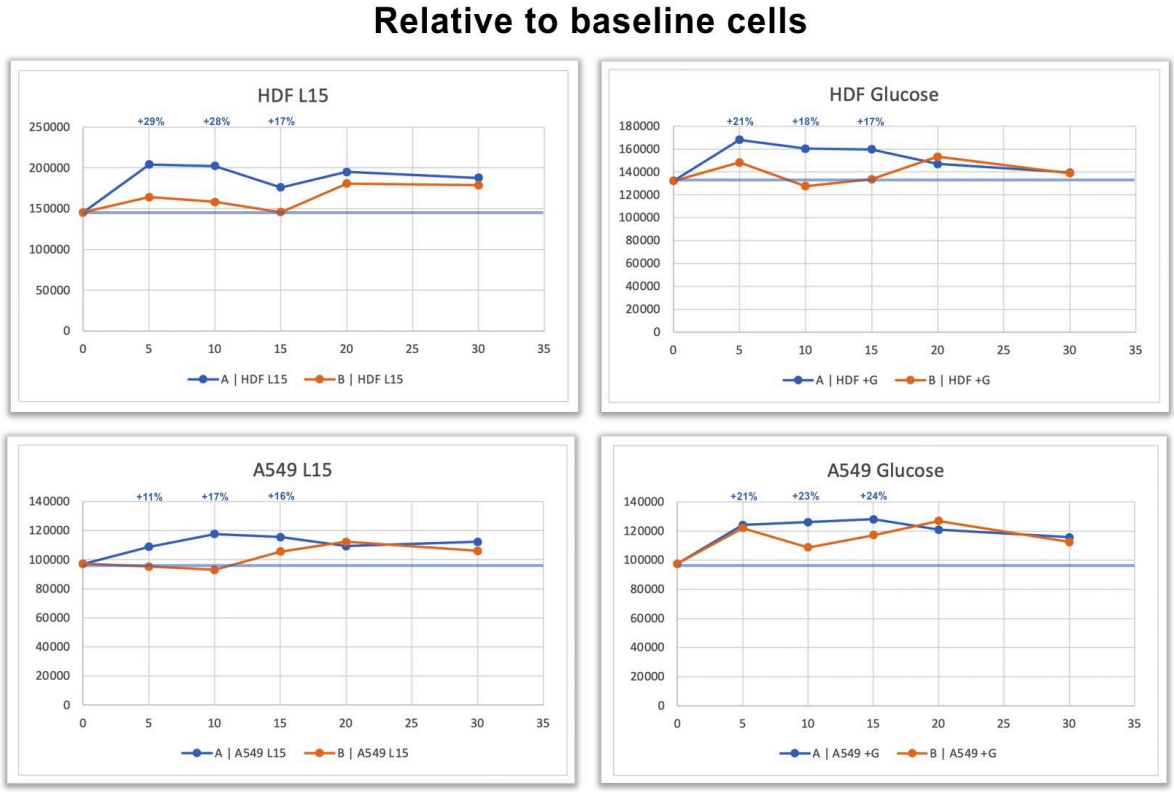

Figure 1. ATP production in A549 and HDF cells cultured in L15 medium ± glucose following quantum field exposure. Measurements were performed in quadruplicate (n = 4 per time point).

Discussion

The results of this experiment indicate that exposure of human cells to a remotely applied quantum field generated by Leela Quantum Tech technology was associated with a measurable, transient increase in ATP production. Both A549 and HDF cell lines demonstrated this response under carefully controlled, double-blind conditions, suggesting that the phenomenon is reproducible across metabolically distinct cell types.

The increase in ATP levels observed in treated cells, ranging from approximately 20 to 29 percent, reflects a biologically meaningful modulation of cellular energy metabolism. The fact that similar changes were detected under both glucose-containing and glucose-free

conditions suggests that the effect is not dependent on a specific metabolic pathway. Instead, it may represent a generalized enhancement of cellular energy production or efficiency, potentially involving mitochondrial or enzymatic regulation.

The experiment was designed to minimize confounding factors that could influence ATP measurements. Conducting the study in a non-CO_2 incubator prevented fluctuations in pH caused by repeated removal of plates, which are known to affect cellular metabolic activity. The use of attachment-compatible plates also ensured consistent cell adherence, a critical factor for fibroblast metabolism and reproducible ATP quantification.

The findings suggest that remote quantum field exposure can transiently modulate cellular bioenergetics. Possible explanations could involve modulation of mitochondrial function, transient shifts in membrane potential, or resonance effects influencing enzymatic activity within the oxidative phosphorylation system. These possibilities remain speculative and require targeted investigation using direct mitochondrial and molecular assays.

The double-blind structure of the study, together with consistent replication across two human cell lines, supports the reliability of the observed response. Future work should address these gaps by including larger sample sizes, detailed statistical validation, and independent replication in additional laboratories.

Overall, this experiment provides preliminary evidence that non-local quantum field exposure may influence ATP production in living cells. The results highlight the need for further studies to explore this effect systematically and to identify the underlying biophysical and biochemical mechanisms responsible for the transient enhancement of cellular energy output.

Conclusion

Under rigorously controlled, double-blind laboratory conditions, exposure of human A549 and HDF cells to a remotely generated quantum field produced by Leela Quantum Tech technology was associated with a transient increase in ATP levels. The response was observed in both glucose-containing and glucose-free environments, indicating that the effect was not limited to a specific metabolic pathway. The magnitude of change—approximately 20–29 percent above control levels—suggests a measurable modulation of cellular bioenergetics.

While the exact mechanism remains undetermined, the findings provide preliminary evidence that quantum field exposure may transiently influence energy production in living cells. These results warrant further investigation using expanded sample sizes, detailed statistical evaluation, and independent replication to confirm reproducibility and explore underlying biophysical pathways.

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